# **Original Research**

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# Induction of genetic variability for quantitative traits in horsegram (*Macrotyloma uniflorum*) through irradiation mutagenesis

S. Priyanka<sup>1</sup>, R. Sudhagar<sup>2\*</sup>, C. Vanniarajan<sup>3</sup>, K. Ganesamurthy<sup>1</sup> and J. Souframanien<sup>4</sup>

<sup>1</sup>Centre for Plant Breeding and Genetics, Tamil Nadu Agricultural University, Coimbatore-641 003, India

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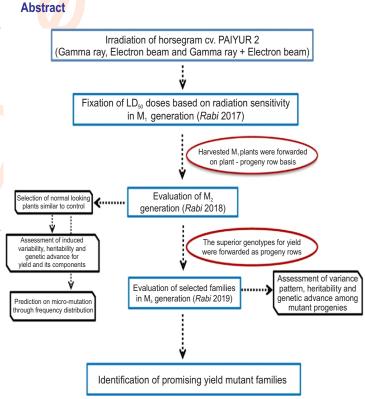
**Aim:** The study was conducted to explore the magnitude of variability induced by gamma rays, electron beam and its combinations for quantitative traits in horsegram (*Macrotyloma uniflorum* (Lam) Verdc.).

**Methodology:** The horsegram cultivar PAIYUR 2 was irradiated with twelve combinations involving G, EB and G+EB. Desirable mutagenic doses were identified in  $M_1$  based on plant injuries and forwarded to M2. Data on 11 yield component traits were recorded on normal looking  $M_2$  plants for identification of micro-mutants. Promising mutants were forwarded to  $M_3$  and the induced variability was ascertained using multivariate statistics.

**Results:** Significant transgressive variants were observed in  $M_2$ . The G+EB combination induced superior variants for yield. A positive shift in mean for yield and its attributing traits was observed in  $M_3$ . Most of the polygenic traits were governed by additive gene effects.

**Interpretation:** The increased mean for most of the polygenic traits in  $M_3$  population proved that selection made on early generation was highly effective. The isolated high yielding mutant progenies of selected families will contribute towards yield improvement in horsegram.

**Key words:** Electron beam, Gamma rays, Horsegram, Induced variability



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<sup>&</sup>lt;sup>2</sup>Sugarcane Research Station, Melalathur, Tamil Nadu Agricultural University, Vellore- 635 806, India

<sup>&</sup>lt;sup>3</sup>Department of Plant Breeding and Genetics, Agricultural College and Research Institu<mark>te, Madurai-625</mark> 104, India

<sup>&</sup>lt;sup>4</sup>Nuclear Agriculture and Biotechnology Division, BARC, Mumbai - 400 085, India

<sup>\*</sup>Corresponding Author Email: sudhagar.r@tnau.ac.in

### Introduction

Horsegram (Macrotyloma uniflorum (Lam) Verdc.) is an autogamous crop under leguminous family with maturity duration of 120 - 180 days (Morris, 2008). It is well adapted to low rainfall areas with average temperature ranging from 18° to 27°C. The seeds are highly nutritious with 16.9 - 30.4% protein content (Patel et al., 1995), 51.9 - 60.9% carbohydrates and 0.58 - 2.06% low lipid content (Bravo et al., 1999; Sodani et al., 2004), lysine (Gopalan, 1989), molybdenum (Bravo et al., 1999), phosphorous, iron, and vitamins viz., carotene, vitamin C, riboflavin, niacin and thiamine (Sodani et al., 2004). It is commonly known as poor man's pulse crop because of its drought tolerant nature with enriched health benefits satisfying the nutritional and economic requirement of marginal farmers. This multipurpose legume is primarily cultivated as a food crop in India, Mauritius, Malaysia, Myanmar, Sri Lanka and Nepal, while as fodder crop in Africa and Australia (Asha et al., 2006). In India, horsegram occupies an area of 3.25 lakh ha with a production and productivity of 1.16 lakh tones and 355 kg ha<sup>-1</sup>, respectively. Southern India is well thought-out as centre of origin for cultivated species of horsegram (Vavilov, 1951; Zohary, 1970).

The crop is highly photo-sensitive with limited productivity that has led to the replacement of horsegram cultivable areas with other crops. Scientific interventions in horsegram would highly facilitate the crop improvement thereby providing nutritional security in the developing countries. Exploitation of variability for yield, crop duration and ideal stature types would provide an opportunity for evolving superior agronomic types. Chahota et al. (2013) reported the existence of limited variability for desirable traits in Indian germplasm accessions. Moreover, hybridization followed by selection remains futile in horsegram because of small floral buds, extreme flower drop and low percentage of pod set (3-4%). Adoption of induced mutagenesis would ease crop improvement with shorter time period (Manjaya and Nandanwar, 2007). Horsegram being a diploid crop (2n=20 and 24), the probability of evolving mutants remains high providing maximum opportunities for selection. Earlier attempts were made on induced mutations in horsegram to explore the effectiveness of different physical (Bolbhat and Dhumal, 2009; Patel et al., 2010) and chemical mutagens (Bolbhat et al., 2012; Kulkarni and Mogle, 2013), but reports on electron beam and its combinations are meagre. Induction of variability through electron beam was found equally effective in relation with gamma rays in legume species (Joshi-Saha et al., 2015; Souframanien et al., 2016). With these views, the experimental study was undertaken to explore the effect of gamma rays (G), electron beam (EB) and combined treatment of gamma rays and electron beam (G+EB) in inducing variability for yield and its related components in horsegram.

# **Materials and Methods**

A well adopted, horsegram cultivar "PAIYUR 2" released from Tamil Nadu Agricultural University (TNAU), India was utilized for inducing of variability through mutagenesis. Irradiations were

carried out at Bhabha Atomic Research Center (BARC), Trombay, Mumbai, India. The mutagen treated seeds viz., 100 Gy, 200 Gy, 300 Gy and 400 Gy of gamma rays, electron beam and G + EB were sown in a randomized block design with two replications, which constituted  $M_1$  population. Assessment of plant injuries at  $M_1$  generation a paved way in determining suitable doses for variability induction.

Each M<sub>1</sub> plant from 200 Gy and 300 Gy of gamma rays; 100 Gy and 200 Gy of electron beam and 100 Gy of G + EB was harvested individually and forwarded to M<sub>2</sub> generation following plant to progeny row method during 2018. Data on 12 biometrical traits viz., days to first flowering, days to maturity, plant height (cm), pod length (cm), number of primary branches per plant, number of pods per cluster, number of clusters per plant, number of pods per plant, number of seeds per pod, hundred seed weight (g), biological yield (g) and single plant yield (g) were recorded for randomly selected 150 normal looking plants similar to control (PAIYUR 2) in each M, population. The selected individual genotypes on seed yield basis were forwarded to M<sub>3</sub> generation as progeny rows during 2019. Similar biometrical observations were recorded among the progenies of selected mutant families in Ma generation. All the mutant generations were raised at experimental plot of Department of Pulses, TNAU during Rabi season.

Commercial package of practices was adopted with a spacial pattern of 30 × 15 cm in all generations. High order statistics viz., skewness and kurtosis (Snedecor and Cochran, 1967) were performed in  $M_2$  population for each mutagen to determine the magnitude of induced micro-mutations. The biometrical data of  $M_2$  and  $M_3$  population were utilized to estimate mean, range, genotypic coefficient of variation (GCV), phenotypic coefficient of variation (PCV) (Burton, 1952), broad sense heritability (H²) (Lush, 1940), genetic advance as percent of mean (GAM) (Johnson  $et\ al.$ , 1955) which provide information on nature and magnitude of variability present in the segregating population and the extent of trait improvement over selection. The estimation of first, second and high order statistics for segregating population are as follows:

First and second order statistics: The variance obtained from the control was taken as error variance ( $\sigma^2 e$ ). The occurrence of variance in mutant population was considered as phenotypic variance ( $\sigma^2 p$ ). The genotypic variance ( $\sigma^2 g$ ) was given by the difference between phenotypic and error variance.

Genotypic Coefficient of Variation (GCV)= $\sqrt{(\sigma^2 g)}$ /Mean × 100 Phenotypic Coefficient of Variation (PCV)= $\sqrt{(\sigma^2 p)}$ / Mean × 100 Heritability (B.S) (H²) =  $(\sigma^2 g/\sigma^2 p)$  × 100

Genetic Advance (GA) =  $(\sigma^2 g/\sigma^2 p) \times k \times \sqrt{(\sigma^2 p)}$  (k = selection differential)

**High order statistics:** The frequency distribution pattern was assessed in  $M_2$  population by the estimates of skewness ( $\beta_1$ ) and kurtosis ( $\beta_2$ ) using SPSS software.

Skewness = 
$$\frac{(n\sum (Y-\mu)^3)}{(n-1)(n-2)\sigma^3}$$

Kurtosis = 
$$\frac{n(n + 1) \sum (Y - \mu)^4}{(n - 1) (n - 2) (n - 3)\sigma^4} - \frac{3(n - 1)^2}{(n - 2) (n - 3)}$$

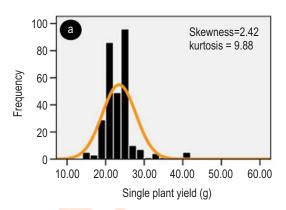
n is the number of observations; Y is the individual observation;  $\mu$  is the sample mean and  $\sigma$  is the standard deviation.

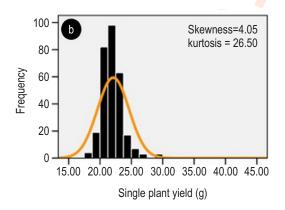
### **Results and Discussion**

Improvement of yield and its related components were found promising in many legume species through induced mutagenesis. Datir (2016) reported the effectiveness of conventional mutagens (gamma rays, ethyl methane sulphonate, sodium azide and combinations) in generation of variants for polygenic traits in horsegram. The mutation induction through

electron beam has proved to be promising because of its high absorbed dosage rate than gamma rays and other radiations (Zhu *et al.*, 2008). The high dosage rate results in increased double stranded DNA breaks leading to induction of several mutations in a population. Therefore, the present study was undertaken to explore the extent of variability induced by gamma rays, electron beam and its combinations in horsegram.

Statistical analyses on M<sub>2</sub> population would be highly beneficial in predicting the induction of micro-mutations (Scossiroli, 1977; Oladosu *et al.*, 2016). Most of the mutant population of M<sub>2</sub> generation exhibited low average mean compared to control for all quantitative traits, except days to first flowering, days to maturity and plant height (Table 1). The shift in mean towards negative direction resulted due to the deleterious effect of employed mutagens on yield component traits. The prolonged duration in flowering and maturity among mutant population has been reported earlier by Rudraswami *et al.* (2006) in horsegram and Manjaya and Nandanwar (2007) in soybean.





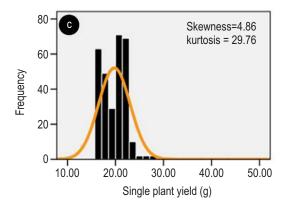


Fig. 1: Frequency distribution pattern of M<sub>2</sub> generation for single plant yield in the irradiated population of: (a) gamma rays; (b) electron beam and (c) gamma rays+electron beam.

 $\textbf{Table 1:} Mean, Shift in mean, range and variance for quantitative traits in $M_2$ population of horsegram cv. PAIYUR 2 and the second property of the secon$ 

Traits	Treatment	Mean	Shift in mean	Range	Variance	Traits	Treatment	Mean	Shift in mean	Range	Variance
	CONTROL	51.17 ± 0.13		54.00 - 49.00	2.38		CONTROL	47.41 ± 0.23		58.00 - 44.00	7.73
	G: 200 Gy	$53.16 \pm 0.21$	1.99	59.00 - 50.00	6.77		G: 200 Gy	$46.30 \pm 0.47$	- 1.11	55.00 - 36.00	32.60
Days to	G: 300 Gy	$52.40 \pm 0.23$	1.23	59.00 - 43.00	7.99	Number of	G: 300 Gy	$43.71 \pm 0.54$	- 3.70	66.00 - 28.00	43.22
first	EB: 100 Gy	$53.46 \pm 0.21$	2.29	61.00 - 47.00	6.63	clusters	EB: 100 Gy	$41.97 \pm 0.75$	- 5.44	91.00 - 31.00	83.23
flowering	EB: 200 Gy	$53.83 \pm 0.21$	5.66	61.00 - 45.00	6.77	per plant	EB: 200 Gy	$38.85 \pm 0.49$	- 8.56	61.00 - 21.00	37.38
•	G+EB: 100 Gy	$51.17 \pm 0.17$	0.00	61.00 - 47.00	4.30		G+EB: 100 Gy	$31.14 \pm 0.54$	- 16.27	54.00 - 24.00	43.65
	CONTROL	$105.60 \pm 0.15$		110.00 - 102.00	3.38		CONTROL	$111.47 \pm 0.51$		124.00 - 105.00	38.97
	G: 200 Gy	$108.23 \pm 0.32$	2.63	115.00 - 103.00	14.80		G: 200 Gy	$70.85 \pm 1.09$	- 40.62	105.00 - 52.00	177.86
Days to	G: 300 Gy	$106.64 \pm 0.27$	1.04	116.00 - 92.00	10.51	Number of	G: 300 Gy	$69.30 \pm 1.24$	- 42.17	142.00 - 42.00	230.82
maturity	EB: 100 Gy	$107.74 \pm 0.32$	2.14	120.00 - 101.00	14.89	pods per	EB: 100 Gy	$84.72 \pm 1.01$	- 26.74	157.00 - 34.00	152.12
	EB: 200 Gy	107.09 ± 0.30	1.49	120.00 - 94.00	13.62	plant	EB: 200 Gy	$68.55 \pm 1.09$	- 42.92	142.00 - 38.00	177.78
	G+EB: 100 Gy	$108.44 \pm 0.33$	2.84	114.00 - 100.00	16.02		G+EB: 100 Gy	$52.01 \pm 1.14$	- 59.46	162.00 - 28.00	194.36
	CONTROL	$72.29 \pm 0.37$	•	80.90 - 64.10	20.46		CONTROL	$5.32 \pm 0.02$		5.70 - 4.68	60.0
	G: 200 Gy	$83.17 \pm 0.85$	10.88	102.30 - 60.20	108.34		G: 200 Gy	$5.25 \pm 0.04$	- 0.07	6.00 - 4.33	0.22
Plant	G: 300 Gy	$87.00 \pm 0.65$	14.71	118.20 - 48.60	62.12	Number of	G: 300 Gy	$5.43 \pm 0.04$	0.11	6.45 - 4.18	0.27
height (cm)	EB: 100 Gy	$74.40 \pm 0.89$	2.11	140.30 - 53.20	117.42	seeds per	EB: 100 Gy	$5.26 \pm 0.04$	90.0 -	6.00 - 4.00	0.21
	EB: 200 Gy	$91.22 \pm 1.11$	18.93	161.20 - 51.30	182.88	pod	EB: 200 Gy	$5.63 \pm 0.05$	0.31	6.67 - 4.33	0.35
	G+EB: 100 Gy	$76.84 \pm 1.05$	4.55	165.20 - 44.50	163.02		G+EB: 100 Gy	$5.59 \pm 0.05$	0.26	6.67 - 4.00	0.32
	CONTROL	$5.12 \pm 0.02$		5.51 - 4.50	0.05		CONTROL	$3.94 \pm 0.01$		4.11 - 3.81	0.01
	G: 200 Gy	$4.57 \pm 0.04$	- 0.55	5.63 - 3.18	0.25		G: 200 Gy	$3.43 \pm 0.02$	- 0.50	4.54 - 3.06	0.04
Pod	G: 300 Gy	$4.65 \pm 0.03$	- 0.47	6.12 - 4.00	0.15	Hundred	G: 300 Gy	$3.68 \pm 0.02$	- 0.26	4.15 - 3.11	90.0
length (cm)	EB: 100 Gy	$4.74 \pm 0.04$	- 0.37	5.70 - 3.40	0.25	seed	EB: 100 Gy	$3.80 \pm 0.01$	- 0.13	4.49 - 3.11	0.03
	EB: 200 Gy	$4.80 \pm 0.03$	- 0.31	5.73 - 3.70	0.13	weight (g)	EB: 200 Gy	$3.88 \pm 0.02$	- 0.06	4.52 - 3.44	0.04
	G+EB: 100 Gy	$4.81 \pm 0.04$	- 0.31	5.77 - 3.18	0.20		G+EB: 100 Gy	$3.74 \pm 0.01$	- 0.20	4.15 - 3.32	0.02
	CONTROL	$8.62 \pm 0.04$		10.00 - 7.00	0.26		CONTROL	$64.62 \pm 0.24$	-	69.00 - 61.00	8.93
	G: 200 Gy	$7.24 \pm 0.08$	- 1.38	11.00 - 5.00	1.06		G: 200 Gy	$65.49 \pm 0.43$	0.87	82.00 - 55.00	27.63
Number of	G: 300 Gy	$7.40 \pm 0.10$	- 1.23	12.00 - 5.00	1.49	Biological	G: 300 Gy	$62.62 \pm 0.55$	- 2.00	112.00 - 51.00	45.38
primary	EB: 100 Gy	$7.93 \pm 0.08$	- 0.69	10.00 - 6.00	0.94	yield (g)	EB: 100 Gy	$66.70 \pm 0.73$	2.08	125.00 - 36.00	78.21
branches	EB: 200 Gy	$7.02 \pm 0.07$	- 1.60	11.00 - 4.00	0.79		EB: 200 Gy	$60.69 \pm 0.52$	- 3.93	70.00 - 47.00	39.60
per plant	G+EB: 100 Gy	$6.68 \pm 0.08$	- 1.95	10.00 - 4.00	96.0		G+EB: 100 Gy	$62.86 \pm 0.50$	- 1.77	95.00 - 56.00	37.59
	CONTROL	$2.83 \pm 0.01$		2.92 - 2.51	0.02		CONTROL	$27.86 \pm 0.10$		28.91 - 25.21	1.40
	G: 200 Gy	$2.25 \pm 0.02$	- 0.59	2.99 - 1.67	0.04		G: 200 Gy	$23.52 \pm 0.16$	- 4.34	36.63 - 20.40	3.58
Number of	G: 300 Gy	$2.01 \pm 0.02$	- 0.82	2.85 - 1.52	0.04	Single plant	G: 300 Gy	$23.59 \pm 0.47$	- 4.27	52.32 - 14.22	34.35
pods per	EB: 100 Gy	$2.13 \pm 0.03$	- 0.70	3.67 - 1.67	0.11	yield (g)	EB: 100 Gy	$22.86 \pm 0.23$	- 5.00	42.86 - 18.53	7.50
cluster	EB: 200 Gy	+1	- 1.07	2.67 - 1.33	0.04		EB: 200 Gy	$21.36 \pm 0.16$	- 6.50		4.00
	G+EB: 100 Gy	$2.09 \pm 0.02$	- 0.74	3.41 - 1.67	0.07		G+EB: 100 Gy	$18.19 \pm 0.29$	- 9.68	47.13 - 16.31	12.42

G - Gamma rays; EB - Electron beam and SE - Standard Error

Table 2: Estimates of frequency distribution pattern in M, population of horsegram cv. PAIYUR 2

Traits	Treatments	Skewness	Kurtosis	Traits	Mutagen	Skewness	Kurtosis
	G	- 0.21	0.23		G	0.59*	-0.48
Days to first flowering	EB	- 0.44*	0.41	Number of clusters	EB	2.14*	10.40*
	G + EB	- 0.49*	2.48*	per plant	G + EB	0.61*	-0.09
	G	0.38*	0.32		G	0.58*	1.06*
Days to maturity	EB	0.89*	2.17*	Number of pods	EB	0.51*	5.19*
•	G + EB	0.35	- 1.47*	per plant	G + EB	3.42*	25.10*
	G	- 0.18	0.94*		G	0.10	-0.89*
Plant height (cm)	EB	1.54*	5.42*	Number of seeds	EB	-0.07	-0.51
• ( )	G + EB	3.71*	20.23*	per pod	G + EB	-0.47*	-0.18
	G	0.02	0.14	' '	G	0.48*	0.41
Pod length (cm)	EB	- 0.30*	0.39	Hundred seed	EB	0.18	2.79*
<b>5</b> ( )	G + EB	- 0.72*	0.74	weight (g)	G + EB	0.23	2.07*
	G	0.51*	1.21*	0 (0)	G	2.59*	15.60*
Number of primary	EB	0.17	0.26	Biological yield (g)	EB	1.86*	15.73*
branches per plant	G + EB	0.99*	0.91*	(3)	G + EB	1.97*	5.05*
	G	0.47*	1.36*		G	2.42*	9.88*
	EB	1.24*	2.39*	Single plant yield (g)	EB	4.05*	26.50*
	G + EB	0.83*	3.86*	(9)	G + EB	4.86*	29.76*

G - Gamma rays; EB - Electron beam; \* Significant at 0.05%

Both positive (G: 300 Gy; EB: 200 Gy and G + EB: 100 Gy) and negative shift (G: 200 Gy and EB: 100 Gy) in mean value were noticed for number of seeds per pod among mutagenic treatments of PAIYUR 2. A decline in number of seeds per pod was observed earlier by Bolbhat and Dhumal (2012) in gamma rays and EMS treated population of horsegram. A similar trend was registered for single plant yield in all mutant population of PAIYUR 2 with the highest reduction at G + EB combination (-9.68 g). A decline in yield may be attributed to frequent occurrence of deleterious alleles in a population. However, several superior mutants were identified in all treatments with increased yield range than control. All the mutagenic treatments exhibited wide range of variation as compared to control (Table 1) indicating the occurrence of micro-mutants in a positive and negative direction for the studied traits. High order statistics viz., skewness ( $\beta_1$ ) and kurtosis (β<sub>2</sub>) provide a better insight in predicting the magnitude and direction of micro-mutations in a population (Misra et al., 2008; Kar et al., 2019). Most of the mutant population exhibited significant skewness and kurtosis indicating the potentiality of employed mutagens in induction of variants (Table 2).

The positive skewed distribution (G: 2.42, EB: 4.05 and G+EB: 4.86) with leptokurtic curve (G: 9.88, EB: 26.50 and G+EB: 29.76) was registered for single plant yield in all irradiated treatments. It implies the occurrence of micro-mutants towards reduction in trait value with presence of extremities over the control range in a population (Fig. 1). Similarly, a leptokurtic curve with skewed pattern towards right was registered in all treated population for biological yield, number of pods per plant, number of pods per cluster and plant height. A sharp curve signifies the presence of outliers or extreme variants in the population providing scope for concerned trait improvement.

Adoption of intense selection in mutant population will result in genetic gain for these traits (Roy, 2000).

The electron beam irradiated population exhibited nonsignificant skewness and kurtosis for number of primary branches per plant ( $\beta_1$ : 0.17 and  $\beta_2$ : 0.26) and number of seeds per pod ( $\beta_1$ : -0.07 and β<sub>2</sub>: -0.51) indicating random occurrence of micromutants at both directions. Balint et al. (1968) reported equal distribution of micro-mutants in either direction for most of the polygenic traits. The induction of micro-mutants with increased trait value was noticed in electron beam (β<sub>1</sub>: -0.44) and its combination (β<sub>1</sub>: -0.49) treatment for days to first flowering. A flattened curve than normal (Platykurtic) was exhibited by G + EB (β<sub>2</sub>: -1.47) mutant population indicating less frequency of superior mutants for maturing duration. The occurrence of variants with increased value for pod length was noticed in electron beam (β<sub>1</sub>: -0.30) and its combination with gamma rays ( $\beta_1$ : -0.72). From the estimates, it is evident that mutagens were potential in inducing extreme variant types for most of the yield components. The occurrence of transgressive variants was analyzed to determine the scope of isolating mutants with improvement in yield component traits (Misra et al., 2008).

The estimates provide clear picture on the extent of variation induced by the mutagens in comparison with control (Table 3). The genotype of M<sub>2</sub> population possessing value higher than the highest trait value of control was considered as positive transgressive variant (PTV) and the trait value lower than the lowest value of control as negative transgressive variant (NTV). The average and maximum transgressive variation were worked out for the PTV's and NTV's to identify the extent of deviations from the control. The negative transgressive variants were

 $\textbf{Table 3}. Frequency and magnitude of transgressive variants in $M_2$ population of horsegram cv. PAIYUR 2 and transgressive variants in $M_2$ population of horsegram cv. PAIYUR 2 and transgressive variants in $M_2$ population of horsegram cv. PAIYUR 2 and transgressive variants in $M_2$ population of horsegram cv. PAIYUR 2 and transgressive variants in $M_2$ population of horsegram cv. PAIYUR 2 and transgressive variants in $M_2$ population of horsegram cv. PAIYUR 2 and transgressive variants in $M_2$ population of horsegram cv. PAIYUR 2 and transgressive variants in $M_2$ population of horsegram cv. PAIYUR 3 and transgressive variants in $M_2$ population of horsegram cv. PAIYUR 3 and transgressive variants in $M_2$ population of horsegram cv. PAIYUR 3 and transgressive variants in $M_2$ population of horsegram cv. PAIYUR 3 and transgressive variants in $M_2$ population of horsegram cv. PAIYUR 3 and transgressive variants in $M_2$ population of horsegram cv. PAIYUR 3 and transgressive variants in $M_2$ population of horsegram cv. PAIYUR 3 and transgressive variants in $M_2$ population of horsegram cv. PAIYUR 3 and transgressive variants in $M_2$ population of horsegram cv. PAIYUR 3 and transgressive variants in $M_2$ population variants in$ 

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Traits	Treatment	NTVF	NTVM	NTVA	MNTV	Traits	Treatment	PTVF	PTVM	PTVA	MPTV
	G: 200 Gy	00					G: 200 Gy	00			
Days to first	G: 300 Gy	02	44.00	2.00	00.9	Number of	G: 300 Gy	90	60.75	2.75	8.00
flowering	EB: 100 Gy	02	47.00	2.00	2.00	clusters	EB: 100 Gy	04	80.00	22.00	33.00
	EB: 200 Gy	10	45.00	4.00	4.00	per plant	EB: 200 Gy	10	61.00	3.00	3.00
	G+EB: 100 Gy	90	47.50	1.50	2.00		G+EB: 100 Gy	00			
	G: 200 Gy	00					G: 200 Gy	00			
Days to	G: 300 Gy	01	92.00	10.00	10.00	Number of	G: 300 Gy	10	142.00	18.00	18.00
maturity	EB: 100 Gy	01	101.00	1.00	1.00	pods per	EB: 100 Gy	02	152.50	28.50	33.00
	EB: 200 Gy	90	100.00	2.00	8.00	plant	EB: 200 Gy	05	136.50	12.50	18.00
	G+EB: 100 Gy	02	100.50	1.50	2.00		G+EB: 100 Gy	02	159.67	35.67	38.00
	G: 200 Gy	80	61.14	2.96	3.90		G: 200 Gy	23	00.9	0.30	0.30
Plant height	G: 300 Gy	10	48.60	15.50	15.50	Number of	G: 300 Gy	31	6.13	0.43	0.75
(cm)	EB: 100 Gy	90	60.17	3.93	10.90	seeds	EB: 100 Gy	12	00.9	0.30	0.30
	EB: 200 Gy	02	51.71	12.39	12.78	per pod	EB: 200 Gy	41	6.23	0.53	0.97
	G+EB: 100 Gy	03	52.80	11.30	19.60		G+EB: 100 Gy	44	6.19	0.49	0.97
Traits	Treatment	PTVF	PTVM	PTVA	MPTV	Traits	Treatment	PTVF	PTVM	PTVA	MPTV
	G: 200 Gy	05	5.63	0.12	0.12		G: 200 Gy	01	4.54	0.43	0.43
Pod length	G: 300 Gy	02	5.90	0.39	0.61	Hundred seed	G: 300 Gy	13	4.13	0.02	0.04
(cm)	EB: 100 Gy	60	2.60	0.09	0.19	weight (g)	EB: 100 Gy	02	4.41	0.30	0.38
	EB: 200 Gy	03	5.73	0.22	0.22		EB: 200 Gy	19	4.22	0.11	0.41
	G+EB: 100 Gy	40	5.59	0.08	0.26		G+EB: 100 Gy	04	4.15	0.04	0.04
	G: 200 Gy	01	11.00	1.00	1.00		G: 200 Gy	38	71.11	2.11	13.00
Number of	G: 300 Gy	01	12.00	2.00	2.00	Biological	G: 300 Gy	03	102.00	33.00	43.00
primary	EB: 100 Gy	00			,	yield (g)	EB: 100 Gy	19	81.60	12.60	26.00
branches	EB: 200 Gy	10	11.00	1.00	1.00		EB: 200 Gy	02	70.00	1.00	1.00
per plant	G+EB: 100 Gy	00					G+EB: 100 Gy	23	74.83	5.83	26.00
	G: 200 Gy	02	2.99	0.07	0.07		G: 200 Gy	02	32.92	4.01	7.72
Number of	G: 300 Gy	00			,		G: 300 Gy	80	37.32	8.41	23.41
pods per	EB: 100 Gy	02	3.50	0.58	0.75	Single plant	EB: 100 Gy	90	32.73	3.82	13.95
cluster	EB: 200 Gy	00			,	yield (g)	EB: 200 Gy	02	39.74	10.83	13.08
	G+EB: 100 Gy	10	3.41	0.49	0.49		G+EB: 100 Gy	10	41.81	12.90	18.22

G - Gamma ray; EB - Electron beam; PTVF - Frequency of positive transgressive variants; PTVA - Average positive transgressive variation; PTVF - Frequency of negative transgressive variants; NTVA - Average negative transgressive variation and MNTV - Maximum negative transgressive variation

**Table 4:** Identified desirable doses for the induction of transgressive variants in horsegram cv. PAIYUR 2

Characters	Mutagenic doses
Days to first flowering Days to maturity Plant height (cm) Pod length (cm) Number of primary branches	G (300 Gy) EB (200 Gy) G+EB (100 Gy) G (300 Gy) and EB (200 Gy)
per plant Number of pods per cluster Number of clusters per plant Number of pods per plant Number of seeds per pod Hundred seed weight (g) Biological yield (g) Single plant yield (g)	EB (100 Gy) EB (100 Gy) EB (100 Gy) and G+EB (100 Gy) EB (200 Gy) and G+EB (100 Gy) EB (100 Gy) EB (100 Gy) G (300 Gy), EB (200 Gy) and G+EB (100 Gy)

G - Gamma rays; EB - Electron beam

considered for days to first flowering and days to maturity in this study. The effective doses were selected on the basis of high occurrence coupled with high average and maximum variation from control. The identified promising doses for extreme variants serve as a component in planning for improving of polygenic traits in horsegram (Table 4). Among the mutagens, G + EB: 100 Gy followed by G: 300 Gy induced high frequency (10 and 08) of superior types with high average (12.90 and 8.41) and maximum transgressive variation (18.22 and 23.41) for single plant yield. These mutagenic doses can be employed in future mutation breeding programmes to achieve extreme variants with enhanced yield (> 8 g) over control. The high frequency (06) of early duration types was noticed in 200 Gy electron beam irradiated population exhibiting variation range of 2 (NTVA) to 8 (MNTV) days over control. A wide variation was noticed among mutagenic treatments for occurrence of transgressive types. Among the mutagens, lower dose of electron beam (100 Gy) were found promising in inducing extreme variants for most of the studied yield component traits.

The estimates of mean  $\pm$  standard error, shift in mean, range and variance of M<sub>3</sub> population are presented in Table 5. The shift in mean towards positive and negative direction was observed in mutagenic treatments which contributed towards increased variance over control. Among mutagens, combination of G + EB exhibited the highest shift in mean value for single plant yield (11.57 g), number of clusters per plant (38.33) and number of pods per plant (17.40). The increased plant height (146.37 cm) coupled with biological yield (121.26 g) in 300 Gy of gamma rays irradiated population seems to be useful for evolving mutant lines with superior fodder value. The extreme range than control was noticed for most of the yield component traits in mutant progenies of M<sub>3</sub> population. Implementation of selection on mutant progenies would assist in identifying promising lines with improved agronomic value. The mean of

single plant yield increased from  $M_2$  to  $M_3$  generation with induction of superior mutants in all irradiated treatments. Imposing direct selection on yield basis in  $M_2$  generation could be a reason for positive shift in mean value (Khan and Qureshi, 2006). The extreme mutants yielding more than 70 g (G: 300 Gy, EB: 200 Gy and G + EB: 100 Gy) were isolated in  $M_3$  generation, which provide scope for yield enhancement in horsegram. A positive shift in mean was observed at all mutagenic treatments of  $M_2$  and  $M_3$  generations for plant height. The increase in mean value over control could be attributed to frequent induction of positive mutations for the trait (Khan and Wani, 2006).

The mean of hundred seed weight, number of clusters per plant, number of pods per plant and biological yield had shifted to positive direction in M<sub>3</sub> generation. A parallel report of positive shift in mean value for polygenic traits was given by Khan et al. (2004). Contrarily, decreased mean value was registered for number of primary branches per plant in both generations (M<sub>2</sub> and M<sub>2</sub>) when compared against control. The induction of mutations with detrimental effects could be reason for reduced mean values in treated population (Singh et al., 2000). The mean of days to first flowering and maturity declined in M<sub>3</sub> population compared to control with few exceptions. Mutants with reduced duration would escape from yield loss due to terminal drought as horsegram is cultivated with residual moisture after the end of North-East monsoon in Southern India. Similar early maturing genotypes were reported by Nilahayati et al. (2019) in gamma irradiated population of soybean. The shift in mean at both positive and negative directions was noticed in mutant population compared to control for number of seeds per pod. Similar report on quantitative traits was given by Mensah and Obadoni (2007) in groundnut. The extent of variability, heritability at broad sense (H2) and genetic advance as percent of mean (GAM) were calculated for M<sub>2</sub> (Table 6) and M<sub>3</sub> populations (Table 7).

Low to moderate variability existed for most of the yield component traits in M<sub>2</sub> generation with few exceptions. The low extent of PCV and GCV was registered for days to first flowering and maturity at both generations. In general, the amount of variability increased in M<sub>3</sub> generation for most of the yield component traits in comparison with M<sub>2</sub> indicating the release of inherent variability. Traits viz., single plant yield, biological yield, number of clusters per plant, number of pods per plant, number of pods per cluster and number of primary branches per plant exhibited high values of PCV and GCV in M<sub>3</sub> generation. Increase in variability estimate provides scope for realizing yield improvement through effective selection (Muduli and Misra, 2008). The estimates of heritability and genetic advance are highly useful in predicting the selection response on quantitative traits. In this study, the estimates of H<sup>2</sup> and GAM tend to increase in M<sub>3</sub> generation for most of the yield component traits with few exceptions. The enhanced heritability in subsequent generation could also be an indicative feature of increased homozygosity of genes for the concerned trait (Wani, 2011). The high H<sup>2</sup> coupled with high GAM indicates the governance of additive gene effects

Table 5: Mean, Shift in mean, range and variance for quantitative traits in  $M_3$  generation of horsegram cv. PAIYUR 2

				)							
Traits	Treatment	Mean	Shift in	Range	Variance	Traits	Treatment	Mean	Shift in	Range	Variance
			mean						mean		
	CONTROL	$52.11 \pm 0.63$		55.00 - 50.00	3.21		CONTROL	$52.11 \pm 1.16$		58.00 - 46.00	10.77
	G: 200 Gy	$51.11 \pm 0.52$	- 1.00	55.00 - 48.00	4.54		G: 200 Gy	$53.00 \pm 2.52$	0.89	69.00 - 31.00	108.00
Days to	G: 300 Gy	$49.98 \pm 0.31$	- 2.13	52.00 - 45.00	4.74	Number of	G: 300 Gy	$83.16 \pm 1.85$	31.05	151.00 - 51.00	167.81
first	EB: 100 Gy	$52.02 \pm 0.36$	- 0.09	60.00 - 44.00	6.50	clusters	EB: 100 Gy	$53.44 \pm 2.82$	1.33	125.00 - 31.00	390.17
flowering	EB: 200 Gy	$52.10 \pm 0.41$	- 0.01	61.00 - 47.00	8.13	per plant	EB: 200 Gy	$58.18 \pm 3.23$	6.07	156.00 - 28.00	510.39
	G+EB: 100 Gy	$51.56 \pm 0.30$	- 0.55	58.00 - 49.00	4.37		G+EB: 100 Gy	$90.44 \pm 3.05$	38.33	182.00 - 45.00	454.85
	CONTROL	$106.11 \pm 0.81$		112.00 - 105.00	5.21		CONTROL	$119.59 \pm 2.73$		138.00 - 108.00	59.62
	G: 200 Gy	$106.78 \pm 0.62$	0.67	111.00 - 102.00	6.62		G: 200 Gy	$121.22 \pm 2.16$	1.63	129.00 - 87.00	79.17
Days to	G: 300 Gy	$106.44 \pm 0.59$	0.33	116.00 - 101.00	16.81	Number	G: 300 Gy	$128.69 \pm 3.85$	60'6	281.00 - 98.00	726.02
maturity	EB: 100 Gy	$104.44 \pm 0.49$	- 1.67	123.00 - 99.00	11.85	of pods	EB: 100 Gy	$119.94 \pm 2.30$	0.35	179.00 - 81.00	260.06
	EB: 200 Gy	$106.00 \pm 0.49$	-0.11	115.00 - 95.00	12.00	per plant	EB: 200 Gy	$119.80 \pm 3.78$	0.20	261.00 - 60.00	699.46
	G+EB: 100 Gy	$108.26 \pm 0.51$	2.15	116.00 - 100.00	12.67		G+EB: 100 Gy	$136.99 \pm 3.92$	17.40	242.00 - 112.00	753.78
	CONTROL	$95.03 \pm 1.58$		101.20 - 88.60	19.99		CONTROL	$5.18 \pm 0.13$		5.55 - 4.67	0.14
	G: 200 Gy	$97.29 \pm 3.15$	2.26	120.60 - 88.60	168.70		G: 200 Gy	$4.70 \pm 0.14$	- 0.48	6.10 - 4.10	0.31
Plant	G: 300 Gy	$146.37 \pm 2.45$	51.33	208.60 - 110.30	294.15	Number of	G: 300 Gy	$4.98 \pm 0.08$	- 0.20	6.00 - 4.11	0.28
height	EB: 100 Gy	$127.39 \pm 2.67$	32.35	169.40 - 62.50	349.00	seeds	EB: 100 Gy	$5.07 \pm 0.07$	- 0.12	6.67 - 4.00	0.23
(cm)	EB: 200 Gy	$120.49 \pm 2.90$	25.45	189.20 - 87.10	411.15	per pod	EB: 200 Gy	$5.60 \pm 0.08$	0.42	6.67 - 4.33	0.32
	G+EB: 100 Gy	$132.77 \pm 3.72$	37.73	220.50 - 76.40	678.51		G+EB: 100 Gy	$5.59 \pm 0.06$	0.40	6.67 - 4.67	0.19
	CONTROL	$4.82 \pm 0.12$		5.40 - 4.40	0.11		CONTROL	$3.46 \pm 0.07$		3.81 - 3.21	0.03
	G: 200 Gy	$4.91 \pm 0.16$	0.09	5.80 - 3.80	0.44		G: 200 Gy	$3.51 \pm 0.05$	0.04	3.83 - 3.20	0.05
Pod	G: 300 Gy	$4.76 \pm 0.09$	- 0.06	6.50 - 3.99	0.41	Hundred	G: 300 Gy	$3.55 \pm 0.03$	60.0	4.12 - 3.25	90.0
length	EB: 100 Gy	$4.78 \pm 0.07$	- 0.04	6.11 - 3.25	0.23	seed	EB: 100 Gy	$3.70 \pm 0.04$	0.24	4.60 - 3.25	0.07
(cm)	EB: 200 Gy	$4.94 \pm 0.09$	0.12	6.30 - 3.50	0.38	weight (g)	EB: 200 Gy	$3.91 \pm 0.05$	0.44	4.80 - 3.10	0.12
	G+EB: 100 Gy	$4.67 \pm 0.08$	- 0.15	6.30 - 3.14	0.32		G+EB: 100 Gy	$3.75 \pm 0.06$	0.28	4.80 - 3.20	0.20
	CONTROL	$8.22 \pm 0.22$		10.00 - 8.00	0.40		CONTROL	$67.22 \pm 0.68$		71.00 - 64.00	3.73
	G: 200 Gy	$7.56 \pm 0.33$	- 0.67	10.00 - 5.00	1.80		G: 200 Gy	$70.33 \pm 1.27$	3.11	82.00 - 64.00	27.33
Number of		$7.02 \pm 0.29$	- 1.20	14.00 - 4.00	4.18	Biological	G: 300 Gy	$121.26 \pm 2.65$	54.04	148.00 - 91.00	344.31
primary	EB: 100 Gy	$6.38 \pm 0.27$	- 28.	14.00 - 4.00	3.64	yield (g)	EB: 100 Gy	$73.82 \pm 3.28$	09.9	222.00 - 51.00	526.07
branches	EB: 200 Gy	$6.38 \pm 0.26$	- 28.	12.00 - 4.00	3.32		EB: 200 Gy	$70.20 \pm 3.61$	2.98	209.00 - 33.00	637.76
per plant	G+EB: 100 Gy	$6.40 \pm 0.24$	- 1.82	12.00 - 4.00	2.80		G+EB: 100 Gy	$96.62 \pm 3.44$	29.40	189.00 - 49.00	579.88
	CONTROL	$2.34 \pm 0.05$		2.52 - 2.00	0.02		CONTROL	$24.56 \pm 0.72$		29.61 - 23.12	4.15
	G: 200 Gy	$2.23 \pm 0.13$	- 0.11	2.87 - 1.62	0.31		G: 200 Gy	$25.94 \pm 1.25$	1.38	35.69 - 15.60	26.65
Number of		$2.55 \pm 0.07$	0.21	3.67 - 1.33	0.27	Single	G: 300 Gy	$33.08 \pm 2.00$	8.52	74.64 - 14.36	195.99
pods per	EB: 100 Gy	$2.15 \pm 0.09$	- 0.19	3.33 - 1.33	0.37	plant	EB: 100 Gy	$27.36 \pm 1.32$	2.81	52.89 - 12.90	85.43
cluster	EB: 200 Gy	$2.48 \pm 0.09$	0.14	3.67 - 1.33	0.38	yield (g)	EB: 200 Gy	+1	3.09	72.74 - 13.40	160.37
	G+EB: 100 Gy	$2.38 \pm 0.09$	0.04	3.33 - 1.33	0.40		G+EB: 100 Gy	$36.13 \pm 1.97$	11.57	75.34 - 15.57	189.85

G-Gamma rays; EB-Electron beam and SE-Standard Error

**Table 6**: Estimates of variability parameters, heritability and genetic advance in M<sub>2</sub> population of horsegram cv. PAIYUR2

				1 - 17							
Traits	Treatment	PCV (%)	GCV (%)	$H^2$ (%)	GAM (%)	Traits	Treatment	PCV (%)	GCV (%)	$H^{2}$ (%)	GAM (%)
	G: 200 Gy	4.53	3.64	64.32	6.01		G: 200 Gy	12.30	10.73	76.14	19.29
Days to	G: 300 Gy	4.74	3.87	99.99	6.51	Number	G: 300 Gy	15.05	13.64	82.13	25.47
first	EB: 100 Gy	4.47	3.57	63.70	2.87	of clusters	EB: 100 Gy	21.80	20.77	90.76	40.75
flowering	EB: 200 Gy	4.49	3.61	64.51	5.97	per plant	EB: 200 Gy	15.52	13.77	78.72	25.17
	G+EB: 100 Gy	3.77	2.52	44.66	3.47		G+EB: 100 Gy	21.28	19.32	82.39	36.12
	G: 200 Gy	3.57	3.13	77.21	2.67		G: 200 Gy	18.85	16.66	90.82	30.32
Days to	G: 300 Gy	3.05	2.51	67.87	4.26	Number	G: 300 Gy	20.24	18.11	80.09	33.39
maturity	EB: 100 Gy	3.59	3.16	77.35	5.72	of pods	EB: 100 Gy	14.60	12.60	74.42	22.39
	EB: 200 Gy	3.46	3.00	75.25	5.36	per plant	EB: 200 Gy	19.48	17.20	78.04	31.31
	G+EB: 100 Gy	3.68	3.26	78.69	2.97		G+EB: 100 Gy	26.87	24.02	79.95	44.25
	G: 200 Gy	12.47	11.23	81.02	20.82		G: 200 Gy	8.91	6.81	58.34	10.71
Plant	G: 300 Gy	80.6	7.45	67.30	12.59	Number	G: 300 Gy	9.55	7.76	60.99	13.00
height	EB: 100 Gy	14.61	13.29	82.72	24.90	of seeds	EB: 100 Gy	8.78	6.65	57.30	10.37
(cm)	EB: 200 Gy	14.87	14.02	88.90	27.23	per pod	EB: 200 Gy	10.54	9.07	74.10	16.09
	G+EB: 100 Gy	16.66	15.59	87.53	30.04		G+EB: 100 Gy	10.08	8.51	71.23	14.79
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	G: 200 Gy	10.79	9.54	78.17	17.38		G: 200 Gy	90.9	5.31	76.91	9.59
Pod	G: 300 Gy	8.33	0.70	64.67	11.10	Hundred	G: 300 Gy	0.70	6.13	83.56	11.54
length	EB: 100 Gy	10.56	9.38	78.88	17.16	seed	EB: 100 Gy	4.75	3.95	69.34	6.78
(cm)	EB: 200 Gy	7.50	5.77	59.19	9.15	weight (g)	EB: 200 Gy	5.18	4.49	75.24	8.02
	G+EB: 100 Gy	9.21	78.7	73.02	13.86		G+EB: 100 Gy	3.42	2.13	38.92	2.74
	G: 200 Gy	14.28	12.41	75.55	22.22		G: 200 Gy	8.04	6.62	67.75	11.22
Number of	G: 300 Gy	16.49	14.97	82.42	28.00	Biological	G: 300 Gy	10.79	89.6	80.41	17.88
primary	EB: 100 Gy	12.28	10.45	72.44	18.32	yield (g)	EB: 100 Gy	13.30	12.52	88.64	24.29
branches	EB: 200 Gy	12.73	10.44	67.25	17.63		EB: 200 Gy	10.40	9.16	77.56	16.62
per plant	G+EB: 100 Gy	14.85	12.72	73.42	22.46		G+EB: 100 Gy	9.77	8.54	76.30	15.36
	G: 200 Gv	8 85	6 97	62 07	11.32		G: 200 Gv	8.06	6.29	88 09	10 10
Minhorad	0.500	1000	0.00	62.07	10.01	Olegio	. 200	27.50	04.0	06.00	10.10
number of	G. 300 Gy EB: 100 Gy	15.37	0.10 11.25	85.09	77.23	olingie plant	G. 300 Gy EB: 100 Gy	24.30 12.02	10.87	93.0Z	40.32 20.15
chister	EB: 200 Gy	11 35	8 96	62.23	14.56	vield (a)	EB: 100 Gy	0.38	7.57	65.73	12.57
i de la composición de la comp	G+EB: 100 Gy	12.04	10.52	76.28	18.93	yield (g)	G+EB: 100 Gy	19.43	18.30	88.75	35.51

PCV (%) - Phenotypic coefficient of variation; GCV (%) - Genotypic coefficient of variation; H² (%) - Heritability at broad sense; GAM (%) - Genetic advance as per cent of mean; G - Gamma rays and EB - Electron beam

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Table 7: Estimates of variability parameters, heritability and genetic advance in M<sub>3</sub> population of horsegram cv. PAIYUR2

Traits	Treatment	PCV (%)	GCV (%)	H² (%)	GAM (%)	Traits	Treatment	PCV (%)	GCV (%)	H² (%)	GAM (%)
	G: 200 Gy	4.17	2.26	29.35	2.52		G: 200 Gy	19.61	18.61	90.03	36.37
Days to first	G: 300 Gy	4.36	2.47	32.27	2.90	Number	G: 300 Gy	15.58	15.07	93.58	30.03
flowering	EB: 100 Gy	4.90	3.49	50.61	5.11	of clusters	EB: 100 Gy	36.96	36.45	97.24	74.04
	EB: 200 Gy	5.47	4.26	60.52	6.82	per plant	EB: 200 Gy	38.83	38.42	97.89	78.30
	G+EB: 100 Gy	4.05	2.09	26.48	2.21		G+EB: 100 Gy	23.58	23.30	97.63	47.43
	G: 200 Gy	2.41	1.11	21.27	1.06		G: 200 Gy	7.34	3.65	24.69	3.73
Days to	G: 300 Gy	3.85	3.20	00.69	5.47	Number	G: 300 Gy	20.94	20.06	91.79	39.59
maturity	EB: 100 Gy	3.30	2.47	56.02	3.80	of pods	EB: 100 Gy	13.45	11.80	77.07	21.35
	EB: 200 Gy	3.27	2.46	56.58	3.81	per plant	EB: 200 Gy	22.08	21.11	91.48	41.60
	G+EB: 100 Gy	3.29	2.52	58.89	3.99		G+EB: 100 Gy	20.04	19.23	92.09	38.02
	G: 200 Gy	13.35	12.53	88.15	24.24		G: 200 Gy	11.93	8.86	55.14	13.55
Plant height	G: 300 Gy	11.72	11.31	93.20	22.50	Number	G: 300 Gy	10.65	7.56	50.37	11.06
(cm)	EB: 100 Gy	14.67	14.24	94.27	28.48	of seeds	EB: 100 Gy	9.48	5.95	39.37	7.69
	EB: 200 Gy	16.83	16.41	95.14	32.98	per pod	EB: 200 Gy	10.02	7.48	55.63	11.49
	G+EB: 100 Gy	19.62	19.33	97.05	39.23		G+EB: 100 Gy	7.88	4.16	27.82	4.52
	G: 200 Gy	13.56	11.82	76.04	21.24		G: 200 Gy	6.24	3.37	29.09	3.74
Pod length	G: 300 Gy	13.52	11.58	73.40	20.44	Hundred	G: 300 Gy	08.9	4.74	48.54	08'9
(cm)	EB: 100 Gy	10.04	7.26	52.23	10.80	seed	EB: 100 Gy	7.11	5.36	26.78	8.32
	EB: 200 Gy	12.45	10.48	70.91	18.18	weight (g)	EB: 200 Gy	8.70	7.48	74.03	13.26
	G+EB: 100 Gy	12.16	9.87	65.91	16.51		G+EB: 100 Gy	12.00	11.07	85.15	21.05
	G: 200 Gy	17.77	15.70	78.08	28.58		G: 200 Gy	7.43	6.91	86.36	13.22
Number of	G: 300 Gy	29.12	27.71	90.55	54.32	Biological	G: 300 Gy	15.30	15.22	98.92	31.18
primary	EB: 100 Gy	29.89	28.22	89.13	54.88	yield (g)	EB: 100 Gy	31.07	30.96	99.29	63.55
branches	EB: 200 Gy	28.54	26.79	88.08	51.79		EB: 200 Gy	35.97	35.87	99.42	73.67
per plant	G+EB: 100 Gy	26.15	24.23	85.89	46.26		G+EB: 100 Gy	24.92	24.84	98.36	51.01
	G: 200 Gy	24.90	24.08	93.50	47.97		G: 200 Gy	19.90	18.29	84.44	34.61
Number of	G: 300 Gy	20.24	19.46	92.47	38.55	Single	G: 300 Gy	42.32	41.87	97.88	85.33
pods per	EB: 100 Gy	28.28	27.50	94.57	55.10	plant	EB: 100 Gy	33.78	32.95	95.14	66.20
cluster	EB: 200 Gy	24.94	24.28	94.74	48.68	yield (g)	EB: 200 Gy	45.81	45.21	97.41	91.92
	G+EB: 100 Gy	26.58	25.90	94.98	52.00		G+EB: 100 Gy	38.14	37.72	97.81	76.84

PCV (%) - Phenotypic coefficient of variation; GCV (%) - Genotypic coefficient of variation; H² (%) - Heritability at broad sense; GAM (%) - Genetic advance as per cent of mean; G - Gamma rays and EB -

for the studied polygenic traits. Similar findings on quantitative traits were reported earlier by Sheeba et al. (2003) and Mensah et al. (2005). Traits viz., days to first flowering and days to maturity exhibited low to moderate H2 coupled with low GAM at both generations indicating preponderance of non - additive gene action. A significant improvement in mean of yield and its component traits was observed with increased H2 and GAM values. It is evident that selection imposed on M2 generation based on single plant yield was highly effective. Similar findings were reported by Solanki and Sharma (2002) in mutant population of lentils. Moreover, variability estimates increased for most of the yield component traits in M3 generation indicating further scope of achieving desirable mutants in advanced generations through selection. In a nutshell, the mutagens used in the study were highly promising for inducing desirable variants in horsegram. The combination of G + EB was found desirable as it exhibited high amount of transgressive variants (PTVF: 10) coupled with improved genetic gain (GAM: 35.51%) in M<sub>3</sub> generation for single plant yield. The stability of isolated high yielding mutant progenies will be test verified in subsequent generation. Thus, the delineated superior mutants offer scope for yield realization in horsegram of Southern India.

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## **Add-on Information**

Authors' contribution: S. Priyanka: Execution, Data compilation, analysis and manuscript preparation; R. Sudhagar: Conceptualization, methodology, execution, data analysis and supervision; C. Vanniarajan: Data execution, review and editing, fund acquisition; K. Ganesamurthy: Supervision, editing and resources and J. Souframanien: Methodology, review, editing, fund acquisition.

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#### References

- Asha, K.I., M. Latha, Z. Abraham, P.K. Jayan, M.C. Nair and S.K. Mishra: Horsegram in India. In: Genetic Resources (Eds.: D. Kumar). Scientific Publisher, Jodhpur, India, p. 11-28 (2006).
- Balint, A., D. Dudits and J. Sutka: Study of quantitative characters of pea in a mutant population. *Novenytermeles*, **17**, 187-196 (1968).
- Bolbhat, S.N. and K.N. Dhumal: Effect of mutagens on quantitative characters in M2 and M3 generation of horsegram (*Macrotyloma uniflorum* (Lam.) Verdc). *Int. J. Sci. Res.*, **2**, 1-7 (2012).
- Bolbhat, S.N. and K.N. Dhumal: Induced macromutations in horsegram [Macrotyloma uniflorum (Lam) Verdc]. Legume Res., 32, 278-281 (2009).
- Bolbhat, S.N., V.D. Bhoge and K.N. Dhumal: Induced mutations in horsegram [Macrotyloma uniflorum (LAM) VERDC)]: Chlorophyll mutations, mutagenic efficiency and effectiveness. Int. J. Life Sci. Pharma. Res., 2, 159-168 (2012).
- Bravo, L., P. Siddhuraju and F. Saura-Calixto: Composition of underexploited Indian pulses. Comparison with common legumes. *Food Chem.*, **64**, 185-192 (1999).
- Burton, G.W.: Quantitative inheritance in grass. In: Proc. of 6<sup>th</sup> International Grassland Congress, p. 24-83 (1952).
- Chahota, R.K., T.R. Sharma, S.K. Sharma, N. Kumar and J.C. Rana: Horsegram. In: Genetic and Genomic Resources of Grain Legume Improvement, Elsevier Inc. Chapters, pp. 293 (2013).
- Datir, S.S.: Genetic improvement in horsegram [Macrotyloma uniflorum (lam) verdc.] (syn. Dolichos biflorus L.) through induced mutations.

  J. Food Legumes, 29, 174-179 (2016).
- Gopalan, C., B.V. Ramashastri and S.C. Balasubramanyan: Nutritive value of Indian foods. National Institute of Nutrition, ICMR, Hyderabad, India (1989).
- Johnson, H.W., H.F. Robinson and R.E. Comstock: Estimates of genetic and environment variability in soybean. *Agronomy J.*, **47**, 314-318 (1955).
- Joshi-Saha, A., K.S. Reddy, V.C. Petwal and J. Dwivedi: Identification of novel mutants through electron beam and gamma irradiation in chickpea (*Cicer arietinum* L.). J. Food Legumes, 28, 1-6 (2015).
- Kar, R.K., T.K. Mishra and B. Pradhan: Studies on frequency distribution, skewness and kurtosis in F<sub>1</sub>M<sub>1</sub> mutant populations of sesame. *Int. J. Curr. Microbiol. App. Sci.*, 8, 1755-1760 (2019).
- Khan, M.R. and A.S. Qureshi: Induced genetic variability in quantitative traits of kabuli chickpea (*Cicer arietinum* L.). *Proc. Pakistan Acad. Sci.*, **43**, 87-94 (2006).
- Khan, S., M.R. Wani and K. Parveen: Induced genetic variability for quantitative traits in *Vigna radiata* (L.) Wilczek. *Pakistan J. Bot.*, 36, 845-850 (2004).
- Khan, S. and M.R. Wani: Induced mutations for yield contributing traits in greengram. *Int. J. Agri. Bio.*, **8**, 528-530 (2006).
- Kulkarni, G.B. and U.P. Mogle: Effects of mutagen on chlorophyll mutation in horsegram [Macrotyloma uniflorum (Lam) Verdcourt]. Biosci. Discov., 4, 214-219 (2013).
- Lush, J.L.: Intra-sire correlations and regression of offspring on dams as a method of estimating heritability of characters. *Proc. American Soci. Ani. Prod.*, **33**, 293-301(1940).
- Manjaya, J.G. and R.S. Nandanwar: Genetic improvement of soybean variety JS 80-21 through induced mutations. *Plant Mut. Rep.*, 1, 36-40 (2007).
- Mensah, J.K. and B. Obadoni: Effect of sodium azide on yield parameters of groundnut (*Arachis hypogea* L.). *African J. Biotech.*, **6**, 668-671 (2007).

- Mensah, J.K., P.A. Akomeah and E.O. Ekpekurede: Gamma induced variation of yield parameters in cowpea (*Vigna unguiculata* (L.) Walp). *Global J. Pure. App. Sci.*, **11**, 327-330 (2005).
- Misra, R.C., P.K. Sahu, C.R. Jali, H.P. Mishra and L.D. Misra: Studies on skewness, kurtosis and transgressive variation in M2 populations of ricebean (*Vigna umbelata*) varieties. *Legume Res. An. Int. J.*, **31**, 94-99 (2008).
- Morris, J.B.: *Macrotyloma axillare* and *M. uniflorum*: Descriptor analysis, anthocyanin indexes, and potential uses. *Genet. Res. Crop Evol.*, **55**, 5-8 (2008).
- Muduli, K.C. and R.C. Misra: Induced polygenic variability in M2 generation and its relationship with production of high yielding mutants in finger millet. *Indian J. Genet.*, **68**, 419-425 (2008).
- Nilahayati, R., D.S. Hanafiah and F. Harahap: The genotype selection of M<sub>3</sub> generation of Kipas Putih soybean with gamma-rays irradiation on agronomic characters, early maturity and high yielding mutants. *Bulg. J. Agric. Sci.*, **25**, 97-102 (2019).
- Oladosu, Y., M.Y. Rafii, N. Abdullah, G. Hussin, A. Ramli, H.A. Rahim, G. Miah and M. Usman: Principle and application of plant mutagenesis in crop improvement: A review. *Biotech. Biotech. Equip.*, **30**, 1-16 (2016).
- Patel, D.P., B.S. Dabas, R.S. Sapra and S. Mandal: Evaluation of horsegram [Macrotyloma uniflorum (Lam.)] germplasm. Nat. Bureau Plant Genet. Res., pp. 37-53 (1995).
- Patel, N.B., N.B.S. Tikka and J.B. Patel: Effect of gamma rays on different parameters under laboratory and field conditions in M1 generation in horsegram. *J. Food Legumes*, **23**, 14-20 (2010).
- Roy, D.: Plant Breeding The analysis and exploitation of variability. Narosa Publishing House, New Delhi, India (2000).
- Rudraswami, P., K.P. Vishwanatha and C. Gireesh: Mutation studies in horsegram (*Macrotyloma uniflorum* (Lam.) Verdc). Proc., LSS-2006, BARC, Mumbai, India, pp. 88-89 (2006).

- Scossiroli, R.E.: Mutations in characters with continuous variation. In: Manual on Mutation Breeding (Tech. Rep. Serial No. 19). IAEA, Vienna, pp. 118-123 (1977).
- Sheeba, G., S.M. Ibrahim, P. Yogameenakshi and S. Babu: Effect of mutagens on quantitative traits in M2 generation in sesame (Sesamum indicum L.). Indian J. Genet., 63, 173-174 (2003).
- Singh, V.P., M. Singh and J.P. Lal: Gamma rays and EMS induced genetic variability for quantitative traits in urdbean (*Vigna mungo* (L.) Hepper). *Indian J. Genet.*, **60**, 89-96 (2000).
- Snedecor, G.W. and W.G. Cochran: Statistical Methods. 6<sup>th</sup> Edn., Oxford and IBH Publishing Co. Pvt. Ltd., New Delhi, India, p. 593 (1967).
- Sodani, S.N., R.V. Paliwal and L.K. Jain: Phenotypic stability for seed yield in rainfed horsegram (*Macrotyloma uniflorum* (Lam.) Verdc.). In: National Symposium on Arid Legumes for Sustainable Agriculture and Trade, pp. 5-7 (2004).
- Solanki, I.S. and B. Sharma: Induced polygenic variability in different groups of mutagenic damage in lentil (*Lens culinaris* Medik.). *Indian J. Genet.*, **62**, 135-139 (2002).
- Souframanien, J., K.S. Reddy, V.C. Petwal and J. Dwivedi: Comparative effectiveness and efficiency of electron beam and 60Co γ-rays in induction of mutations in blackgram [Vigna mungo (L.) Hepper]. J. Food Legu., 29, 1-6 (2016).
- Vavilov, N.I.: Phyto-geographic basis of plant breeding. Chron. Bota., 13, 13–54 (1951).
- Wani, A.A.: Induced polygenic variability for quantitative traits in chickpea var. Pusa-372. *Comuni. Sci.*, **2**, 100-106 (2011).
- Zhu, H., J.Z. Xu, S.Q. Li, X.Y. Sun, S.D. Yao and S.L. Wang: Effects of high energy pulse-electron beam radiation on biomacromolecules. Sci. China Ser. B: Chem., 51, 86-91 (2008).
- Zohary, D.: Centers of diversity and centers of origin. In: Genetic Resources in Plants (Eds.: O.H. Frankel and F. Bennett). Oxford, Blackwell Scientific Publications, pp. 33-42 (1970).